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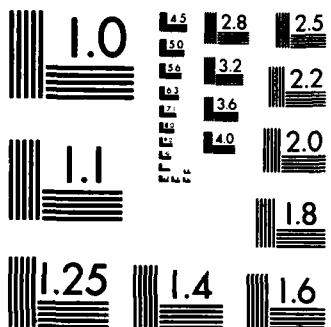
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20. ABSTRACT (Continue on reverse side if necessary and identify by block number) This report reviews progress made under contract DAMD17-79-C-9071. Investigations were carried out on red cells stored in CPDA-2 including: estimations of osmotic fragility of stored RBCs before & after reinfusion into volunteer donors. BAGPM was studied as a preservative medium to improve red cell viability. Storage of platelets made from blood collected in CPD adenine preservatives was studied.		

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BLOOD PRESERVATION STUDY

ANNUAL PROGRESS REPORT

January 1981

E. BEUTLER

Supported by

U.S. ARMY MEDICAL RESEARCH AND DEVELOPMENT COMMAND
FORT DETRICK, FREDERICK, MARYLAND 21701

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1. Study of blood stored in CPD and in modified CPD solutions containing adenine.

1.1. Background

CPD (citrate-phosphate-dextrose) has been routinely used as the standard medium for preservation of blood for many years. Red cells from blood which has been collected in CPD can only be stored for 21 days. After this, they are no longer sufficiently viable for transfusion. It has been clearly established that the addition of adenine to CPD preservative solution prolongs the length of time that red cells may be stored to 35 or 42 days (1). Moreover, higher concentrations of glucose than those present in CPD are required to sustain red cell concentrates for prolonged periods of time in adenine-containing media (2).

CPD-A1 contains a 1.25 times the glucose content of CPD and sufficient adenine to provide a concentration of 0.25 mM in the blood-preservative mixture. Higher glucose concentrations were not incorporated into this preservative because of the existence of anecdotal reports that high glucose concentrations increased the rate of fall of the pH of platelet concentrates. CPD-A1 was studied by a cooperative group (3) with the demonstration that red cells from whole blood stored for 35 days had viability of $80.53 \pm 6.44\%$ ($\bar{x} \pm 1$ S.D.) while red cell concentrates with a hematocrit averaging 75% had a mean viability of $71.38 \pm 10.3\%$ ($\bar{x} \pm 1$ S.D.). After these studies were concluded, it was discovered that the manufacturer of the blood bags had inadvertently prepared the bags used from an obsolete plastic film, PL 130, rather than the new plastic formulation, PL 146, which had been intended. These studies have now been repeated in PL 146, with results quite similar to those

obtained in PL 130. In both their series of studies, it was observed that most or all of the glucose was exhausted from cells in some units packed at hematocrits of 75 \pm 5% and stored for 35 days. There is reason for concern that under field conditions, where the storage temperature may sometimes rise to higher than 4° and where packing of the red cells, a parameter which is difficult to control, may frequently exceed 80%, that CPD-A1 may prove to be suboptimal for the storage of red blood cells. In the meantime, it has become clear that higher glucose concentrations do not harm platelets. For this reason, two new preservatives, believed to represent improved formulations for storage of cell concentrates have been devised and manufactured by Fenwal Laboratories. These formulations, designated CPD-A2 and CPD-A3 contain respectively 1.75 and 2.00 times the amount of glucose present in CPD and sufficient adenine to provide a concentration of 0.5 mM in the blood-preservative mixture.

Data which have been accumulated over the past 15 years clearly indicate that ATP levels are a disappointing parameter with respect to predicting the viability of stored red cells (4-7). We have also been attempting to identify measurements which may prove to be more useful than ATP in predicting whether or not a given storage system provides red cells which will be viable on reinfusion.

1.2. Progress in current period (January 1980 through December 1980).

1.2.1. Biochemical studies and studies of viability of stored cells.

450 ml of blood has been collected from each of 13 volunteer donors. After an 8-hour holding period at room temperature the red cells were packed to a hematocrit of 80% and stored at 4°C for 42 days without agitation. The first five units were stored in a lying position; the

storage position of the subsequent 8 units was alternated between lying and standing. Viability studies were carried out on 11 of the 13 units. One unit was spuriously reported to be contaminated and was therefore not reinfused, and one donor withdrew her consent to participate in the study. The results of biochemical studies and of the viability of the units is summarized in Table I.

1.2.2. Auxiliary studies.

In the course of these investigations various auxiliary studies were undertaken to aid in better definition of the "storage lesion."

1.2.2.1. Studies of osmotic fragility.

The osmotic fragility of stored red cells was investigated both before and after reinfusion into volunteer donors. The osmotic fragility of red cells stored in CPD-A2 was found to be greatly increased when measured directly by estimating lysis in graded phosphate-buffered salt solutions. Much of this increase in osmotic fragility was found to be due, however, to the accumulation of lactate in the stored cells. Lactate leaves red cells only slowly and therefore exerts a marked osmotic effect. When stored red cells were pre-equilibrated with isotonic sodium chloride solutions until the lactate had been removed from the cells the osmotic fragility was increased to a much smaller extent. However, a "tail" of osmotically fragile cells could still be detected. Studies to determine the fate of these cells were performed by devising a method of sequential hemolysis which permitted us to measure the osmotic fragility of the reinfused cells using the release of ^{51}Cr from the labeled cells as an indicator of hemolysis. These investigations

demonstrated that the osmotic fragility of stored cells rapidly returns to normal after their reinfusion into the circulation. The normalization of the osmotic fragility of reinfused cells does not apparently depend principally upon removal of the most fragile cells, but rather on correction of the increased osmotic fragility of cells in the circulation. It is postulated that this correction is due to transfer of osmotically active substances from the red cell after re-infusion.

1.2.2.2. Studies of complement binding.

According to Dr. I. Szymanski, the binding of complement to red cells may prove to be a useful predictor or viability. In the course of some of the viability studies which have been carried out red cells have been frozen in glycerol according to Dr. Szymanski's instructions and have been shipped to her for study.

1.2.2.3. Studies of deformability

The measurement of erythrocyte deformability by use of the ektocytometer has been pioneered by Mohandas and Bessis. Recently, Mohandas has instituted certain marked improvements in this technology. Samples of blood samples which were studied for viability have been shipped to Dr. Mohandas in San Francisco for measurements of deformability using the ektocytometer.

2. Studies with BAGPM

2.1 Background

BAGPM is a bicarbonate-containing preservative medium designed in this laboratory (8,9) for the purpose of maintaining viability and

2,3-DPG levels of packed red cells in a superior fashion. Presumably because of the removal of platelets, granulocytes and fibrinogen, the system minimizes formation of microaggregates (8). The excellent 2,3-DPG preservation observed depends upon buffering of the stored cells with bicarbonate with loss of CO₂ from the system. This can be accomplished either by using containers made from a very permeable plastic film or by introducing into the bags an efficient CO₂ absorbing system. Our investigations suggest that calcium hydroxide, either in small sialastic ("baggies") or embedded in sialastic blocks ("blockies"), might be suitable internal CO₂ absorbing devices.

In earlier studies it was demonstrated that Ca(OH)₂ added to blood produced marked hemolysis. Hence, leakage of calcium hydroxide from the sialastic membrane "baggie" constituted a serious potential hazard. Therefore, the use of calcium hydroxide embedded sialastic blockies of 3 x 3 x 1 cm size were studied as replacements of the old baggies.

Investigations with varying amounts of calcium hydroxide showed that 6 g calcium hydroxide was the most satisfactory amount to provide adequate preservation of 2,3-DPG without compromising the levels of ATP. It was observed initially that calcium hydroxide shed from the raw surfaces when stored at room temperature in ACD and to some extent also in CPD. These difficulties were accentuated after autoclaving in ACD. They were overcome by providing a coating of liquid silicone rubber over the surface of the "blockie."

Recently, Högman and his colleagues have introduced a solution for the storage of red cells, designated SAG (i.e., saline-adenine-glucose) (10). The use of this solution would seem to us to be a retrogressive step (11) because it does little more than to provide additional glucose

to the cells and to reduce the viscosity of the cell suspension. BAGPM, in contrast, provides excellent maintenance of 2,3-DPG levels.

2.2 Studies Performed (January 1980 through December 1980)

Unfortunately, no further studies of BAGPM have been carried out in the current contract period. We have been in continued touch with Fenwal Laboratories but have been unable to obtain from them suitable packs containing CPD and a calcium hydroxide impregnated "blockie" in the primary pack and satellites containing BAGPM. Various technical obstacles have been overcome by Fenwal but their progress in providing the needed materials to us has been painfully slow.

3. Studies of platelet storage.

3.1 Background

The transfusion of platelets has become an important part of the clinical practice of medicine. Platelets, unlike red cells, can only be maintained in storage for up to 72 hours. Little is known about the factors which limit the storageability of platelets. Although there are a number of relatively minor changes in the level enzymatic activities of metabolic intermediates in stored platelets (12,13) none of these seem to decline as abruptly at 72 hours as does the viability of reinfused platelets. Of all of the parameters of platelet storage which have been investigated, pH appears to correlate most closely with post-storage viability (14). In our studies of platelet preservation in CPD solutions containing various amounts of glucose we observed that there was a correlation between the rate of fall of pH and the number of residual white cells in the platelet preparations (15).

The purpose of the studies described below was to more accurately appraise the effect of leukocyte contamination on platelet storage and to determine whether practical means could be devised to maintain platelet pH by removing white cells from the platelet preparations.

3.2 Progress in the current period (January 1980 through December 1980)

Extensive investigations have been carried out to determine the best conditions for removal of leukocytes from platelets during storage and to measure the effect of leukocyte removal on pH of the stored platelets. It was found that centrifuging platelet rich plasma at 500 g for 5 minutes resulted in removal of 80 to 90% of the leukocytes with loss of 10 to 20% of the platelets. Centrifuging at appreciably higher speeds or for longer periods of time resulted in greater platelet loss while lower centrifuge speeds or shorter times of centrifugation resulted in less complete leukocyte removal (Table II). The fall in pH and lactate production of platelet concentrates prepared from such treated and control (untreated) plasmas were investigated (see Tables III and IV, V). Although the fall in pH appeared to be less when leukocyte numbers were decreased, the differences observed were relatively slight and it was felt that the loss of platelets and extra labor involved in preparing leukocyte-poor platelet preparations probably did not represent an acceptable means for prolonging platelet storage.

4. Publications

The following publications supported, in part, by this contract, appeared during 1980:

Beutler E, West C: An Improved Assay and some Properties of Phosphoglycolate Phosphatase. *Anal Biochem.* 106:163-168, 1980.

Beutler E, Kuhl W: Guanosine Triphosphatase Activity in Human Erythrocyte Membranes. *Biochim Biophys Acta*, 601:372-379, 1980.

Beutler E, Kuhl W: Platelet Glycolysis in Platelet Storage: IV. The Effect of Supplemental Glucose and Adenine. *Transfusion*, 20:97-100, 1980.

TABLE I - RESULTS OF BIOCHEMICAL AND RE-INFUSION STUDIES
PERFORMED ON BLOOD COLLECTED FROM 13 DONORS AND STORED IN CPD-A₂

Storage Position (L = Lying (S = Standing))	Donor	NB	CC	WC	JS	MG	LP	DR	PG	AM	RP	PN	KL	TB	Day	
		24 hr Viability (%)	42	93.6	90.1	81.4	90.4	87.9	-	-	78.9	75.1	85.4	79.2	81.6	75.6
Hematocrit (%)	0 42	35 80	39 78	35 80	39 81	36 80	34 84.5	39 80.3	29 80	33.5 81	37 85	34 80	41 80	41 81	43 80.8	
Glucose (mg %)	0 42	549 97	589 137	587 148	599 127	590 115	553 95	517 76	531 124	598 79	526 127	534 129	566 146	564 146	564 153	
ATP (μ Moles per gm Hb)	0 42	5.03 3.61	4.05 3.71	3.80 2.38	4.60 3.22	4.20 3.27	4.71 3.56	4.48 3.62	3.87 1.68	4.16 2.07	4.89 3.71	3.86 2.55	4.49 2.96	4.40 2.30		
2,3 DPG (μ Moles per gm Hb)	0 42	14.9 0.3	11.2 0.2	12.8 0.1	14.6 0.3	11.6 0.3	12.5 0.1	14.3 0.1	11.1 0.2	10.0 0.1	13.0 0.1	13.7 0.5	10.2 0.2	14.0 0.2	14.0 0.1	
Plasma Hb (mg %)	0 42	14 420	15 268	17 720	19 860	10 910	12 2263	26 264	10 613	7 1875	11 2775	10 2263	10 1750	20 1750	21 1568	
pH (4°C)	0 42	7.57 6.81	7.61 6.92	7.57 6.87	7.63 6.92	7.58 6.91	7.61 6.78	7.51 6.76	7.43 6.82	7.50 6.77	7.74 6.77	7.53 6.79	7.55 6.78	7.62 6.83		
Na ⁺ (mM)	0 42	169 94	174 109	170 97	171 87	174 92	181 77	171 83	166 76	170 80	182 75	172 80	176 81	174 82		
K ⁺ (mM)	0 42	3.3 97	3.1 83	3.4 94	3.3 100	3.3 92	3.9 117	3.6 104	3.2 102	3.3 98	3.3 114	3.6 98	3.3 102	3.3 102		

TABLE II

THE EFFECT OF CONDITIONS OF CENTRIFUGATION OF
PLATELET-RICH PLASMA (PRP) ON LEUKOCYTE AND PLATELET YIELD

WBC	235	300 g;	80	34	305
Platelet count	300×10^3	5 min	210×10^3	70	$1,150 \times 10^3$
WBC	265	400 g;	70	26	270
Platelet count	420×10^3	5 min	330×10^3	78	$1,590 \times 10^3$
WBC	304	500 g;	40	13.1	40
Platelet count	336×10^3	5 min	347×10^3	103	790×10^3
WBC	1215	500 g;	30	2.4	80
Platelet count	400×10^3	5 min	310×10^3	77.5	$1,350 \times 10^3$
WBC	1520	500 g;	30	1.9	60
Platelet count	375×10^3	5 min	334×10^3	89.0	713×10^3
WBC	210	500 g;	30	14.2	125
Platelet count	188×10^3	5 min	202×10^3	107	534×10^3
WBC	395	500 g;	25	6.3	45
Platelet count	352×10^3	5 min	282×10^3	80.1	$1,492 \times 10^3$
WBC	765	600 g;	75	9.8	135
Platelet count	608×10^3	5 min	341×10^3	56.0	$1,280 \times 10^3$
WBC	680	600 g;	15	2.2	60
Platelet count	475×10^3	5 min	270×10^3	56.8	$1,170 \times 10^3$

TABLE III. CHANGES IN LACTATE CONCENTRATION IN STORED PLATELET CONCENTRATES WITH VARYING LEUKOCYTE COUNTS

E; additional centrifugation to remove leukocytes.
C; control, with no extra centrifugation.

Subject	WBC, mm ⁻³	Platelets x 10 ³ /mm ³	Volume (ml) platelet Concentrate	μMoles Lactate/ml platelet concentrate								
				0	1	2	3	4	5	6	7	8
G. H. (E)	60	713	60	1.06	2.39	2.128	4.18	6.689	-	-	-	-
C. S. (C)	200	871	60	1.78	4.57	5.58	7.07	-	-	-	-	-
<u>Day 11</u>												
P. M. (C)	1,310	832	60	1.13	2.34	3.19	4.65	5.75	6.71	12.44	-	12.2
C. S. (E)	125	534	60	.97	1.75	2.39	2.89	3.68	3.69	8.6	-	5.98
C. W. (E)	20	202	25	1.09	2.04	2.92	4.48	5.23	5.56	15.16	-	-
<u>Day 6</u>												
V. W. (C)	1,385	1,386	60	.917	3.83	7.81	9.36	11.76	12.2	17.2	-	21.3
S. Y. (E)	255	538	60	1.077	1.81	2.77	3.28	3.86	3.99	6.6	-	7.47
D. A. (C)	2,575	2,780	25	3.35	6.87	10.85	13.44	16.48	15.06	20.3	-	24.65
S. H. (E)	45	1,498	25	1.96	3.63	5.94	7.28	8.43	12.4	17.95	-	23.58
<u>Day 9</u>												
D. G. (C)	3260	1,334	34.5	1.45	4.09	5.71	7.73	-	11.7	-	14.56	15.61
G. B. (E)	60	1,495	51.85	1.48	3.29	5.02	6.83	-	9.95	-	13.55	14.14
E. C. (C)	2,573	1,162	61.0	1.13	2.5	3.2	4.88	-	7.15	-	10.13	11.12
C. S. (E)	55	1,045	65.7	.665	1.7	2.85	4.45	-	5.97	-	9.23	11.55

TABLE IV. pH CHANGES IN STORED PLATELET CONCENTRATES WITH VARYING LEUKOCYTE COUNTS

E; additional centrifugation to remove leukocytes.
C; control, with no extra centrifugation.

Subject	<u>WBC</u> mm^3	Platelets $\times 10^3/\text{mm}^3$	Concentrate	Volume (ml)				pH, Day			
				0	1	2	3	4	5	6	7
G.H. (E)	60	713	60	7.19	7.18	7.26	7.22	7.23	-	-	7.18
C.S. (C)	200	871	60	7.13	6.94	7.1	7.12	-	-	-	-
P.M. (C)	1,310	832	60	7.12	7.22	7.19	7.34	7.22	7.3	7.34	-
C.S. (E)	125	534	60	7.2	7.25	7.28	7.49	7.39	7.5	7.58	-
C.W. (E)	20	202	25	7.2	7.37	7.42	7.69	7.6	7.7	7.59	-
V.W. (C)	1,385	1,386	60	7.20	7.04	6.93	6.86	6.84	6.76	6.42	6.56
S.Y. (E)	255	538	60	7.28	7.35	7.42	7.44	7.49	7.51	7.58	7.65
D.A. (C)	2,575	2,780	25	7.11	7.0	7.02	6.94	6.95	6.75	6.37	6.25
S.H. (E)	45	1,498	25	7.24	7.22	7.24	7.27	7.21	7.01	6.67	6.54
D.G. (C)	3260	1.334	34.5	7.17	7.16	7.23	7.24	-	7.23	-	7.13
G.B. (E)	60	1,495	51.85	7.19	7.16	7.13	7.10	-	7.08	-	6.99
E.C. (C)	2,573	1,162	61.0	7.2	7.19	7.19	7.19	-	7.2	-	7.12
C.S. (E)	55	1,045	65.7	7.19	7.2	7.22	7.21	-	7.21	-	7.10
										7.09	7.08

CHANGES IN PH AND LACTATE CONCENTRATION IN STORED PLATELET CONCENTRATES WITH VARYING LEUKOCYTE COUNTS

Lactate and pH change in stored platelet concentrates
E; additional centrifugation to remove leukocytes
C; control, with no extra centrifugation.

Subject	<u>WBC</u> mm ⁻³	Platelets x 10 ³ /mm ³	Volume (ml) platelet Concentrate						pH, Day	Day	μmoles Lactate/ml concentrate	
			1	2	3	4	5	6				
G.H. (C)	7,000	1,062	70	7.6	7.06	7.01	7.0	6.88	-	6.91	3.24	2.54
S.W. (E)	250	955	62	7.3	7.2	7.4	7.4	7.3	-	7.2	1.04	1.54
D.A. (C)	7,226	242	48	7.3	6.84	6.36	5.9	5.76	-	5.74	-	18.82
G.B. (C)	1,086	3,133	43	7.2	7.14	6.93	6.93	7.03	-	6.96	-	10.48
S.H. (E)	170	1,156	47	7.27	7.25	7.32	7.33	7.34	-	7.25	-	6.68
L.T. (E)	40	1,545	48	7.23	7.03	7.02	7.03	7.08	-	7.05	-	5.03
S.D. (C)	730	2,667	43	7.17	7.04	6.98	-	-	-	-	2.06	5.88
E.G. (E)	305	1,146	54	7.36	7.41	7.51	-	-	-	-	1.52	2.09
L.S. (E)	270	1,594	48	7.35	7.32	7.36	-	-	-	1.22	1.25	4.36
J.L. (E)	40	790	55	7.36	7.41	7.42	7.49	-	-	7.59	1.52	2.17
D.C. (E)	80	1,350	60	7.29	7.25	7.22	7.31	-	-	7.33	1.38	2.62
D.G. (C)	2,010	1,650	42	7.22	6.96	6.82	6.78	6.78	-	-	-	-
S.H. (E)	135	1,280	50	7.28	7.19	7.17	7.23	7.28	-	-	-	-
E.V.A. (E)	60	1,170	43	7.28	7.27	7.29	7.36	7.41	-	-	-	-

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